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# In Silico-Predicted Cytokeratin Fragment 21-1 (Cyfra 21-1) Immunogenic Epitopes for The Early Detection of Nasopharyngeal Carcinoma

Brian Umbu Rezi Depamede<sup>1</sup>, Bachti Alisjahbana<sup>2</sup>, Sulaiman Ngongu Depamede<sup>3</sup>, Sanny Hafidhoh Siti Nururrohmah<sup>1</sup>, Ilham Tryawan<sup>1</sup>, Muhammad Yusuf<sup>4</sup>

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Abstract: Nasopharyngeal Carcinoma, an aggressive cancer in the head and neck region is mostly diagnosed at an advanced stage and thus has a poor prognosis. Cytokeratin Fragment 21-1 (CYFRA 21-1) is one of the oncomarkers discovered in saliva. For developing a diagnostic kit, CYFRA 21-1 antibodies and immunogens are needed. Natural CYFRA 21-1 immunogen is difficult to obtain, so it must be made in-silico using bioinformatics. We aim to predict CYFRA 21-1 immunogenic epitope to produce antibodies polyclonal against CYFRA 21-1, which can be used to develop NPC diagnostics. The immunogenic epitopes were predicted and chosen based on antigenicity, surface accessibility, and hydrophilicity, then characteristic was analyzed and evaluated. The epitope candidates were compared with other saliva protein biomarkers to find if there were cross-reactions. CYFRA 21-1 consists of 57 amino acids, where two immunogenic epitopes (C3 and D2) were chosen. The Ramachandran Plot of both epitopes shows that 100% of the amino acids were in the favoured area. Epitopes C3 and D2 have no cross-reaction with other protein biomarkers. The predicted immunogenic epitopes have the potential as antigen to produce antibodies for developing saliva-based immunodiagnostics to early diagnose NPC patients.

**Keywords:** antibody, cancer, cytokeratin fragment 21-1, epitope, NPC, saliva

Abstrak: Karsinoma Nasofaring, kanker agresif di daerah kepala dan leher sebagian besar didiagnosis pada stadium lanjut sehingga memiliki prognosis yang buruk. Cytokeratin Fragment 21-1 (CYFRA 21-1) adalah salah satu penanda kanker yang ditemukan dalam saliva. Untuk mengembangkan kit diagnostik, diperlukan antibodi dan imunogen CYFRA 21-1. Imunogen CYFRA 21-1 alami sulit diperoleh, sehingga harus dibuat secara in-silico menggunakan bioinformatika. Kami bertujuan untuk memprediksi epitop imunogenik CYFRA 21-1 untuk menghasilkan antibodi poliklonal terhadap CYFRA 21-1, yang dapat digunakan untuk mengembangkan alat diagnostik Kanker Nasofaring (KNF). Epitop imunogenik diprediksi dan dipilih berdasarkan antigenisitas, aksesibilitas permukaan, dan hidrofilisitas, kemudian dianalisis dan dievaluasi karakteristiknya. Kandidat epitop dibandingkan dengan biomarker protein saliva lainnya untuk menemukan apakah ada reaksi silang. CYFRA 21-1 terdiri dari 57 asam amino, di mana dua epitop imunogenik (C3 dan D2) dipilih. Plot Ramachandran dari kedua epitop tersebut menunjukkan bahwa 100% asam amino berada di area yang disukai. Epitop C3 dan D2 tidak memiliki reaksi silang dengan biomarker protein lainnya. Epitop imunogenik yang diprediksi memiliki potensi sebagai antigen untuk memproduksi antibodi sebagai bahan pengembangan imunodiagnostik berbasis saliva untuk mendiagnosis pasien KNF secara dini.

Kata kunci: antibodi, kanker, cytokeratin fragment 21-1, epitop, KNF, saliva.

#### INTRODUCTION

Nasopharyngeal Carcinoma (NPC) is an aggressive cancer in the head and neck region, endemic in Southeast Asia, South China and North

Africa (Lin *et al.* 2023). NPC has an incidence rate of 4-25 cases per 100,000 people in these endemic areas (50-100 times more than in other regions) (Wong *et al.* 2021). In 2018, there were an estimated 129,079

<sup>&</sup>lt;sup>1</sup>Department of Biotechnology, Graduate School, Universitas Padjadjaran, Jl. Dipati Ukur No. 35, Bandung, 40132, West Java, Indonesia.

<sup>&</sup>lt;sup>2</sup>Department of Internal Medicine, Faculty of Medicine, Universitas Padjadjaran, Hasan Sadikin General Hospital, Bandung, Jl. Pasteur No. 38 Bandung. Kel. Pasteur Kec. Sukajadi, 40161, West Java, Indonesia.

<sup>&</sup>lt;sup>3</sup>Faculty of Animal Science, Mataram University, Jln. Majapahit No. 62 Mataram, NTB, 83125, Indonesia

<sup>&</sup>lt;sup>4</sup>Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Padjadjaran, Jalan Raya Bandung – Sumedang km. 21, Sumedang 45363, West Java, Indonesia.

<sup>\*</sup>Corresponding author: m.yusuf@unpad.ac.id

diagnosed cases of NPC internationally (85% occurred in Asia), with a mortality rate of 72,987. This makes NPC the 23rd-highest cancer incidence and the 21st cause of death in the world. In Southeast Asia alone, NPC ranks 9th for all cancer incidences and 8th for the number of deaths from cancer (Chang et al. 2021). In Indonesia, in 2012, the prevalence of NPC was 6.2 per 100,000 population, with a total of 13,000 new cases (Kadriyan et al. 2022).

NPC is a malignant tumour that originated in the nasopharyngeal epithelial tissue, undergoing a complex multifactor regulatory process, appearing on the posterior and side walls of the nasopharyngeal area (Xu & Lou 2021). The occurrence of NPC is believed to be the result of an interaction between Epstein-Barr virus (EBV) infection, smoking and alcohol consumption, environmental factors, as well as genetics (Tang et al. 2021). Although the standard treatments are surgery, radiation and chemotherapy, the overall prognosis of NCP remains poor (Liu et al. 2021). This is because 80% of NPC cases are newly diagnosed at an advanced stage (stages III & IV). The delay in diagnosis is due to the asymptomatic NPC at an early stage, high metastatic rate, and difficult access for examination of local primary tumours in the structures of the nasal cavity (Siak et al. 2021).

The success of early detection of cancer plays a very important role in getting a better prognosis (Adeoye et al. 2022). However, at this time, there is no gold standard as to which test is best for the early detection of NPC (Liu et al. 2021). The most popular diagnostic test today is a tissue biopsy, but this procedure is invasive, complicated, painful, timeconsuming, and can harm the patient (Patel et al. 2022). For this reason, an accurate non-invasive method is needed for the early detection and monitoring of NPC using specific biomarkers (Zambonin & Aresta 2022). An example of a protein in saliva that has a high potential to be a biomarker in epithelial cell cancer is Cytokeratin fragment 21-1 (CYFRA 21-1) (Tofighi et al. 2021). As a soluble fragment of Cytokeratin 19 (KRT19), CYFRA 21-1 works as a proteolytic (Xu et al., 2022). This biomarker is released in large quantities in saliva, dissolves and is released during the process of cell apoptosis. CYFRA 21-1 levels in normal individuals is around  $3.06 \pm 0.25$  ng/mL, while in cancer individuals it increases to  $17.46 \pm 1.46 \text{ ng/mL}$ (Rajkumar et al. 2015; Jafari & Hasanzadeh 2020).

CYFRA 21-1 is a marker recognized by two monoclonal antibodies against KRT19 fragments in the serum. The two antibodies' epitopes were KS 19.1 and BM 19.21, found to be located within helix 2B of the rod domain of KRT19. For the catcher antibody (KS 19.1), the epitope is located within the amino acid sequences 311-335, and for the detection antibody (BM 19.21), the epitope is located within the amino acid sequences 346-367 (Jose *et al.* 2013). NPC cases can be detected more quickly if there are practical diagnostic tools. One example of a practical

diagnostic tool is the Immunoassay test, which needs antibodies for its development (Michel *et al.* 2020). To make CYFRA 21-1 antibody, the antigen is needed to be used as the vaccine. The problem is that CYFRA 21-1 antigen is hard to obtain naturally. So, it is necessary to use another method, namely with an antigens that uses peptide fragments to get the desired antibody (Toepak *et al.* 2022).

With the development of biotechnology, the usage of epitope vaccines has increased due to insilico technology, which uses bioinformatic tools and databases to produce safe and encouraging results (Toepak et al. 2022). This approach has several advantages, including the ability to reduce the amount of time and money spent, build persistent immunity to the desired response, and eliminate the unwanted immune response by building specialized structures (Parvizpour et al. 2020). The appropriate technique to assess changes in biomarkers in saliva as an initial diagnostic that can produce a clinical impact on NPC disease has not been found (Guruduth et al. 2021). In this study, we aim to identify CYFRA 21-1 immunogenic epitope that can be used to produce antibodies polyclonal against CYFRA 21-1, which later on will be used in the development of NPC non-invasive diagnostics.

#### MATERIALS AND METHODS

### Target protein sequence identification and retrieval

CYFRA 21-1 is part of KRT19, hence the first step of our study is retrieving the protein sequence of KRT19. The Universal Protein Resource (Uni-Prot) database (<a href="https://www.uniprot.org/">https://www.uniprot.org/</a>) provided the FASTA formatted amino acid sequence for the protein KRT19 (Accession no. P08727) (Rezaei *et al.* 2023). The protein sequence of CYFRA 21-1 was chosen based on the specific epitope region starting from KS 19.1 to BM 19.21 (311-367) (Jose *et al.* 2013).

#### **Epitope prediction and selection**

The IEDB server (http://tools.iedb.org/bcell/) was used to predict immunogenic CYFRA 21-1 epitopes. Antigenicity, surface accessibility, and hydrophilicity were factors taken into consideration when choosing the epitope. The antigenicity, surface accessibility, hydrophilicity scores were obtained using the Kolaskar and Tongaonkar antigenicity prediction, Emini surface accessibility prediction, and Parker hydrophilicity prediction methods in the IEDB server. The immunogenicity prediction algorithms' threshold output values were used to determine if a putative immunogenic epitope should be included. prediction Bepipred linier epitope (https://services.healthtech.dtu.dk/services/BepiPre d-2.0/) were also used with the threshold of 0.4 (Guevarra et al. 2020; Rezaei et al. 2023).

#### **Epitope physicochemical characterization**

The Physicochemical characterization of the epitope construct was assessed using the ExPASyProtParam database (http://web.expasy.org/protparam/).

ExPASyProtParam is used to calculate numerous physical and chemical parameters like the number of amino acids and their composition, theoretical isoelectric point (PI), molecular weight, instability index, grand average hydropathicity (GRAVY), extinction coefficient, estimated half-life, atomic composition, and the aliphatic index (Akash *et al.* 2023; Rezaei *et al.* 2023).

## Secondary and tertiary structure evaluation, 3D structure modeling, refinement and validation

Using the GOR4 server (<u>https://npsa-pbil.ibcp.fr/cgi-</u>

bin/npsa\_automat.pl?page=/NPSA/npsa\_gor4.html ), the epitopes' secondary structure was evaluated to predict the percentage of strand, helix and coil. The epitopes' tertiary structure was then modelled the **I-TASSER** using server  $(\underline{https://zhanggroup.org/I-TASSER/}).$ YASARA Energy Minimization server (<u>https://www.yasara.org/minimizationserver.htm</u>) was utilized to refine the best model structure of the epitopes construct. Then, the model structure was analyzed by using Ramachandran plots (Swiss-Model: https://swissmodel.expasy.org/) (Rezaei et al. 2023).

#### Homology and protein structural comparison

We performed sequence alignment of the selected epitope (C3 and D2) with multiple protein biomarkers in saliva to prevent positive crossreaction (Table 1). Pairwise and multiple sequence alignment of the protein sequence was performed using ClustalW. Phylogenetics was analyzed by the ETE3 and the BIONJ option. Both ClustaW and ETE3 were embedded in the Genome.jp website (https://www.genome.jp/toolstools bin/clustalw). The I-TASSER server was also used to find proteins that are structurally similar to the chosen epitope by evaluating the TM-score and identity percentage. TM-score is a metric for evaluating the topological similarity of protein structures. The TM-score ranges from 0 to 1, with 1 denoting a perfect match between two structures. According to stringent PDB statistics, structures with scores below 0.17 correspond to randomly selected unrelated proteins, whilst those with scores above 0.5 are assumed to have the same fold in SCOP/CATH. Uniprot and the bgee.org website was used to analyze the origin of the organism, classification and function of each protein (Sharma & Dubey 2020; Dong et al. 2023; Rezaei et al. 2023).

#### Proteolytic cleavage analysis

Regarding the stability of peptide antigens, it is necessary to carry out proteolytic cleavage analysis. PeptideCutter server (<a href="https://web.expasy.org/peptide cutter/">https://web.expasy.org/peptide cutter/</a>) were used to analyze the frequencies of the amino acids in proteolytic cleavage sites. Out of a total of 38 proteases, PeptideCutter performs a digestion and offers detailed findings, including the locations of the cleavage sites, peptide sequences, lengths, and masses (Maillet 2020; Goh & Hahn 2021).

#### RESULT AND DISCUSSION

The study of biomarkers for NPC has seen significant advancements, one of which is cytokeratin (CK). CYFRA 21-1 is a fragment of KRT19, which is one of the CK proteins that make up the cell cytoskeleton. Although CYFRA 21-1 does not directly influence the development of NPC, it's important to note that NPC is a type of squamous cell carcinoma that forms in the epithelial layer of the nasopharynx, where all epithelial cells express CK protein. CK protein is a helpful indicator for identifying epithelial differentiation. When cancer cells lysis or necrosis, some CK components are released into the body. One of these proteins is which is expressed and can immunohistochemically detected in the cytoplasm of epithelial cancer cells. Therefore, KRT19 can be used as a tumor marker for many malignancies, including NPC. The soluble part of KRT19 that is released into body fluids is known as CYFRA 21-1 (Sulaiman 2022; Adusumilli et al. 2023). Based on numerous studies, CYFRA 21-1 is a promising biomarker for the detection, staging, and monitoring of various cancers using different techniques and samples (Lei et al. 2019; Liu et al. 2019; Rudhart et al. 2020; Tofighi et al. 2021; Zhao et al. 2021).

To detect cancer using CYFRA 21-1 as a biomarker, we need to create antibodies against CYFRA 21-1. To do this, we require a specific antigen. Antigen that is specific for cancer detection have been challenging to produce, and there is no one-size-fits-all method or tool for rational specific antigen production. The process of developing specific antigen requires several steps. By mapping thousands of biological components in silico, computational approaches can dramatically cut down on the time and expense of vaccine development. Specific antigen (vaccine) development and analysis using this method can start with proteome retrieval, epitope prediction, epitope selection, molecular interactions, and immune response simulation (Patronov & Doytchinova 2013; Parvizpour et al. 2020; Prawiningrum et al. 2022; Toepak et al. 2022).

### Target protein sequence identification and retrieval

In this study, the KRT19 protein sequence was retrieved in FASTA format from the Uniprot

database which consists of 400 amino acids. From the KRT19 protein sequence, CYFRA 21-1 protein sequence was identified starting from 311-367 which consists of 57 amino acids. The sequence is shown in Figure 1. Determination of this sequence was based on the results of previous studies that the sequence can be recognized by two monoclonal antibodies, namely KS 19.1 (known as catcher epitope) and BM 19.21 (known as detector epitope) (Jose *et al.* 2013). Based on this, it can be assumed that the specific antigen developed from this sequence can produce catcher and detector antibodies that are equivalent to the monoclonal antibodies developed by Jose *et al.* (2013). To prove this, it was necessary to carry out immunogenicity testing

#### **Epitope prediction and selection**

Initially, in predicting and selecting the catcher and detector epitope we used two approaches. The first approach was to use four servers to predict the immunogenic epitopes. The four servers are IEDB, ABCpred, BCEPS, and BepiPred (Jespersen *et al.* 2017; Ras-Carmona *et al.* 2021; Dar *et al.* 2022; Rezaei *et al.* 2023). Of the four servers, 7 epitopes were obtained which had a length of 16 amino acids. However, after looking at the antigenicity, surface accessibility, and hydrophilicity parameters, the seven epitopes had fewer immunogenic values (data not shown). Our next approach was to predict epitope one by one using the IEDB server (Kolaskar and Tongaonkar for antigenicity prediction, Emini for

surface accessibility prediction, and Parker for hydrophilicity prediction) with the default threshold value (Guevarra *et al.* 2020). Assessment of these three parameters was very important in designing specific antigens because it increases the likelihood of producing antibodies specific to CYFRA 21-1.

In Table 2, six immunogenic sequences were identified. Further testing of the 6 candidates and based on the highest scores, epitopes C3 and D2 were determined (Table 3). When comparing the D2 epitope candidate with the original BM 19.21 epitope, the D2 epitope has higher antigenicity and meets all three immunogenic parameters. Meanwhile, the C3 epitope when compared to the original KS epitope has antigenicity/immunogenicity score. This high immunogenicity score plays an important role in producing antibodies, especially in diagnostic development. Bepipred analysis (IEDB), shows these two epitopes are unique. From the position of the immunogenic peptides (Figure 2), we can assume that both epitopes would be a good candidate for sandwich (one as a catcher and the other as the detection).

#### **Epitope physicochemical characterization**

The next step was to look at the physicochemical characterization of the two selected epitope candidates. The results using the ExPASyProtParam server showed that the epitopes C3 and D2 have a length of 27 amino acids, and have a relatively small

| No. | Protein biomarker                         | Related disease              | Range   |
|-----|---|------------------------------|---|
| 1.  | Interleukin-1β                            | Periodontitis                | 161.51 <sup>(a)</sup> - 1312.75 <sup>(b)</sup> (pg mL <sup>-1</sup> ) |
| 2.  | Interleukin-8                             | Oral squamous cell carcinoma | 210.20 <sup>(a)</sup> - 1718.61 <sup>(b)</sup> (pg mL <sup>-1</sup> ) |
| 3.  | Vascular endothelial growth factor (VEGF) | Oropharyngeal cancer         | $280^{(a)} - 4321^{(b)} (pg mL^{-1})$                                 |
| 4.  | Triosephosphate isomerase (TPI1)          | Gastric Cancer               | $> 800^{(a)} - < 400^{(b)} (U \text{ mL}^{-1})$                       |
| 5.  | α-amylase                                 | Stress                       | <50-100 <sup>(a)</sup> $- >$ 150 <sup>(b)</sup> (U mL <sup>-1</sup> ) |

Table 1. Protein biomarkers in saliva (Dong et al. 2023).

MTSYSYRQSSATSSFGGLGGGSVRFGPGVAFRAPSIHGGSGGRGVSVSSARFVSSSSSGA YGGGYGGVLTASDGLLAGNEKLTMQNLNDRLASYLDKVRALEAANGELEVKIRDWYQKQG PGPSRDYSHYYTTIQDLRDKILGATIENSRIVLQIDNARLAADDFRTKFETEQALRMSVE ADINGLRRVLDELTLARTDLEMQIEGLKEELAYLKKNHEEEISTLRGQVGGQVSVEVDSA PGTDLAKILSDMRSQYEVMAEQNRKDAEAWFTSRTEELNREVAGHTEQLQMSRSEVTDLR RTLQGLEIELQSQLSMKAALEDTLAETEARFGAQLAHIQALISGIEAQLGDVRADSERQN QEYQRLMDIKSRLEQEIATYRSLLEGQEDHYNNLSASKVL

Figure 1. CYFRA 21-1 sequence identification (underlined)

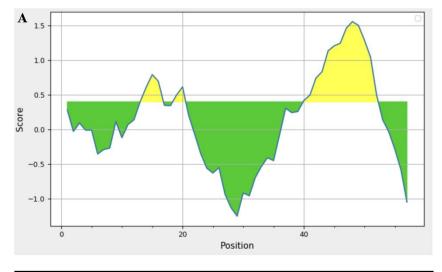
<sup>(</sup>a) Value in normal condition; (b) Value in disease condition.

Table 2. Predicted immunogenic epitopes

| Epitopes |    | Start | Peptide sequence            | Length |
|----------|----|-------|-----------------------------|--------|
| Catcher  | C1 | 311   | QSQLSMKAALEDTLAETEARFGAQLA  | 26     |
|          | C2 | 312   | SQLSMKAALEDTLAETEARFGAQLAH  | 26     |
|          | C3 | 311   | QSQLSMKAALEDTLAETEARFGAQLAH | 27     |
| Detector | D1 | 338   | IQALISGIEAQLGDVRADSERQNQEY  | 26     |
|          | D2 | 338   | IQALISGIEAQLGDVRADSERQNQEYQ | 27     |
|          | D3 | 340   | ALISGIEAQLGDVRADSERQNQEYQRL | 27     |

Table 3. Epitope candidates compared to the original CYFRA 21-1 epitope.

| Epitopes | Antigenicity | Accessibility | Hydrophilicity |
|----------|--------------|---------------|----------------|
|          | (> 1.011)    | (> 1.000)     | (> 2.182)      |
| C1       | 1.011        | 1.309         | 2.031          |
| C2       | 1.014        | 1.029         | 1.881          |
| C3       | 1.014        | 1.573         | 2.033          |
| D1       | 1.013        | 1.889         | 2.519          |
| D2       | 1.013        | 2.888         | 2.648          |
| D3       | 1.011        | 3.843         | 2.537          |
| KS 19.1  | 1.009        | 1.295         | 2.028          |
| BM 19.21 | 0.983        | 8.249         | 3.418          |





**Figure 2.** [A] Immunogenic parts of the CYFRA 21-1 epitope based on Bepipred linier epitope prediction (colored yellow, position 13-16 (TLAE), 19-20 (AR), and 40-52 (GDVRADSERQNQE). [B] 3D visualization of the alpha helix structure of CYFRA 21-1, and the immunogenic parts of CYFRA 21-1 epitopes (Red circles represent oxygen atoms, blue circles represent nitrogen atoms, and dark gray circles represent carbon atoms).

molecular weight of 2917.24 Da and 3032.27 Da respectively. If we assess the instability index, it can be seen that the C3 epitope is more stable than the original KS 19.1 epitope. However, the D2 epitope here was considered an unstable protein compared to the BM 19.21 epitope. In this case, if an epitope is unstable, the solution that can be done is to combine the epitope with another protein such as protein carrier to increase the stability of the protein (Tîrziu *et al.* 2023).

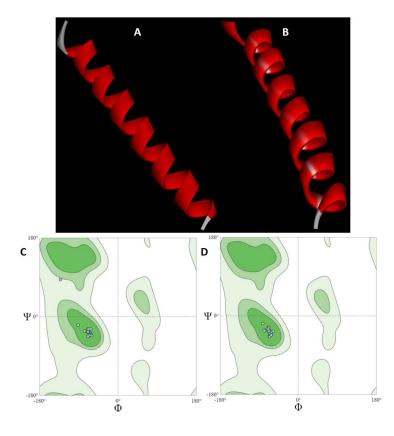
### Secondary & tertiary structure evaluation and 3D structure refinement & validation

The second structure was evaluated via GOR4, and the tertiary structure was built using the I-Tasser server. Epitope C3 secondary evaluation showed a 62.96% helix, 7.41% strand, and 29.63% coil. Whereas for epitope D2 had 55.56% helix, 7.41% strand, and 37.04% coil. From tertiary structure evaluation, the best model of the epitope construct was selected based on the highest C-score (range from -5 to 2). The C-score indicates the confidence score of the protein model evaluated by the I-TASSER server. The best antigen model C-score for C3 was 0.32, with an estimated TM-score 0.76±0.10 and RMSD 1.0±1.0A, whereas the C-score for D2 was 0.30, with an estimated TM-score 0.75±0.10 and RMSD 1.0±1.0A. Both epitopes C3 and D2 had a

good confidence score which means the model has a good quality structure. The best 3D structure model was then refined using YASARA server, which is shown in Figure 3. After being refined using the YASARA server, both epitopes were validated using the Ramachandran Plot. Based on the findings, 100% of the amino acids were in the favoured area for epitopes C3 and D2 (Figure 3).

#### Homology and protein structural comparison

The specific antigen design aims to produce antibodies that can be used to develop a saliva-based diagnostic tool to detect CYFRA 21-1 in NPC patients. For this reason, it is necessary to carry out a comparative analysis between the C3 and D2 epitopes against several biomarker proteins that are commonly found in human saliva. From the sequence alignment (Supplementary Figure 1), the gap open penalty value used in the ClustalW was set to 10.0, the gap extension penalty was set to 0.1, and the weight matrix used was BLOSUM. The minimum pairwise alignment score for both epitope was 11, and the highest score was 14 for epitope C3 and 18 for epitope D2. The homology between epitope C3 and D2 with other protein saliva were analyze using ETE3, and were presented in a phylogenetic tree (Figure 4). From the homology and the overall results of the pairwise alignment score that is below 50%,



**Figure 3.** (A) Refined model of epitope C3; (B) Refined model of epitope D2; (C) C3 Ramachandran Plots; (D) D2 Ramachandran Plots.

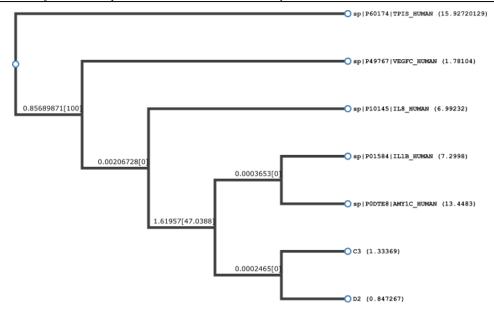


Figure 4. Homology between epitope C3 and D2 with other protein saliva.

| Table 4. | Sequence | alignment | with | other | protein | saliva. |
|----------|----------|-----------|------|-------|---------|---------|
|          |          |           |      |       |         |         |

|   | Pairwise alignment score (%) |            |  |  |  |  |
|---|------------------------------|------------|--|--|--|--|
|   | Epitope C3                   | Epitope D2 |  |  |  |  |
| Interleukin-1β                            | 14                           | 11         |  |  |  |  |
| Interleukin-8                             | 11                           | 14         |  |  |  |  |
| Vascular endothelial growth factor (VEGF) | 14                           | 18         |  |  |  |  |
| Triosephosphate isomerase (TPI1)          | 14                           | 14         |  |  |  |  |
| α-amylase                                 | 14                           | 14         |  |  |  |  |

means there would be minimal positive crossreaction between the epitopes with the other protein biomarker found in saliva (Table 4).

The results of analysis using I-TASSER, it shows that there are proteins that are structurally similar to the C3 and D2 epitopes. Based on Uniprot ID, in both C3 and D2, ten types of proteins were obtained that were structurally similar. The origin of the organism, classification and function of each protein are presented in Table 5 for epitope C3 and Table 6 for epitope D2.

Based on the results of homology analysis and comparison of protein structures using ClustalW, it can be seen that the average pairwise alignment score between epitopes C3 and D2 with other protein biomarkers is below 50%. The analysis results indicate that epitopes C3 and D2 are not closely related to the five biomarker proteins in saliva, so the likelihood of cross-reaction is low. Cross-reaction can occur when epitopes have identical sequences or conformations with the same structure as other proteins or antigens (Mitchell *et al.* 2018). Determining if a cross-reaction has occurred based on sequence similarity is not highly reliable, as proteins form a structure. Assessing cross-reactions based on

structure can enhance accuracy and validity (Buraphaka *et al.* 2024). Therefore, homology between the three-dimensional structures of the two epitopes with other proteins was conducted using the I-TASSER server. Based on the structure, the five biomarker proteins in saliva did not appear in the results of the three-dimensional structure homology analysis from the I-TASSER server.

Further analysis using I-TASSER, we figured out that both epitopes C3 and D2 have some similar structure proteins. We retrieve ten proteins that have the closest structure for each epitope. From this analysis, the origin, TM-score, identity score, classification, function and whether it is found in saliva can be seen. As mentioned in Table 5, of the ten proteins that are structurally similar to the C3 epitope, four are found in the Homo Sapiens species. Meanwhile, from Table 6, it can be seen that there are four proteins structurally similar to the D2 epitope found in human (Homo sapiens) species. The eight proteins found in human (Homo Sapiens) that are similar to the C3 and D2 epitopes can be found in saliva (produced by minor salivary glands). However, if we look at the TM-score and identity score that is quite small, hence the cross-reaction between the

**Table 5.** Protein structurally close to epitope C3

| Rank | Protein PDB Hit  | Uniprot<br>ID | TM-<br>score* | IDEN*<br>(%) | Organism  | Classification      | Function   | Presence<br>in saliva |
|------|--|---------------|---------------|--------------|---|---------------------|--|-----------------------|
| 1    | 7CGPC  (Mitochondrial import inner membrane translocase subunit Tim29) | Q9BSF4        | 0.942         | 0.074        | Homo sapiens  | Translocase         | A part of the TIM22 complex, which facilitates the entry and insertion of multi-pass transmembrane proteins into the inner membrane of the mitochondria. The membrane potential serves as the external driving factor for the twin-pore translocase that the TIM22 complex generates. Functions in the assembly of the TIM22 protein into the TIM22 complex and is necessary for the stability of the TIM22 complex. may interact with TOMM40 to encourage cooperation between TIM22 and TOM complexes (Callegari <i>et al.</i> , 2016). | Yes                   |
| 2    | 6RW8A  (A component of insecticidal toxin complex (Tc))                | D3VHH9        | 0.942         | 0.111        | Xenorhabdus nematophila<br>(strain ATCC 19061 /<br>DSM 3370 / CCUG<br>14189 / LMG 1036 /<br>NCIMB 9965 / AN6) | Toxin               | N/A  | No                    |
| 3    | 3A68C<br>(Ferritin-4,<br>chloroplastic)                                | Q948P5        | 0.939         | 0.111        | Glycine max   | Oxidoreducta-<br>se | Keeps iron in a soluble, safe, and accessible state. Iron is taken up in the ferrous state and deposited as ferric hydroxides after oxidation, which is necessary for iron homeostasis (By similarity) (Masuda <i>et al.</i> 2010).  | No                    |
| 4    | 7O41D<br>(TrwG protein)  | O50335        | 0.939         | 0.037        | Escherichia coli  | Membrane<br>protein | N/A  | No                    |

| 5 | 7T3UA  (Inositol 1,4,5- trisphosphate receptor type 3) | Q14573 | 0.938 | 0.148 | Homo sapiens  | Metal<br>transport    | It is a receptor for inositol 1,4,5-trisphosphate, a second messenger that, by analogy, mediates the release of intracellular calcium and is involved in maintaining the balance of calcium ions in cells (Rönkkö <i>et al.</i> 2020).  | Yes |
|---|--|--------|-------|-------|---|-----------------------|---|-----|
| 6 | 5MRCCC<br>(uL3m)                                       | P31334 | 0.938 | 0.037 | Saccharomyces cerevisiae<br>(strain ATCC 204508 /<br>S288c) | Ribosome              | A part of the mitochondrial ribosome (mitoribosome), a special translational apparatus in charge of producing proteins from the mitochondrial genome, including at least some of the crucial transmembrane components of the mitochondrial respiratory chain. Translation products are cotranslationally integrated into the inner membrane of the mitochondria by the mitoribosomes, which are connected to it (Pfeffer <i>et al.</i> 2015). | No  |
| 7 | 7JGDA1  (Erythrocyte membrane protein 1)               | Q6UDW7 | 0.937 | 0.037 | Plasmodium falciparum                                       | Sugar binding protein | N/A   | No  |
| 8 | 7EU7A  (Glutamate receptor ionotropic, NMDA  1)        | Q05586 | 0.932 | 0.074 | Homo sapiens  | Membrane<br>protein   | It is a part of NMDA receptor complexes, heterotetrameric ion channels that are ligand-gated and have significant calcium permeability as well as voltage-dependent sensitivity to magnesium. Glycine must bind to the zeta subunit of the channel, glutamate must bind to the epsilon subunit, and membrane depolarization is necessary to remove Mg2+-induced channel blockage (Chen <i>et al.</i> 2021).                                   | Yes |
| 9 | 7RPMA  (Neuronal acetylcholine receptor subunit        | P36544 | 0.930 | 0.148 | Homo sapiens  | Membrane<br>protein   | AChR undergoes a significant conformational shift after binding acetylcholine, which affects all subunits and causes the opening of an ion-conducting channel across the plasma membrane. Alpha-bungarotoxin shuts down the channel (Kalashnyk <i>et al.</i>  | Yes |

|    | alpha-7) |       |       |                  |         | 2023). |    |
|----|----------|-------|-------|------------------|---------|--------|----|
| 10 | 6M6ZA    | 0.930 | 0.074 | Escherichia coli | De novo | N/A    | No |
|    | (TMH4C4) |       |       |                  | protein |        |    |

<sup>\*</sup> TM-score: a metric for assessing the topological similarity of protein structures. \* IDEN: the percentage sequence identity in the structurally aligned region.

**Table 6.** Protein structurally close to epitope D2

|   | Protein PDB Hit     | Uniprot | TM-    | IDEN* | Organism          | Classification | Function  | Presence  |
|---|---------------------|---------|--------|-------|-------------------|----------------|---|-----------|
|   |                     | ID      | score* | (%)   |                   |                |   | in saliva |
| 1 | 6SO5C               | O00258  | 0.919  | 0.074 | Homo sapiens      | Membrane       | Required for the endoplasmic reticulum (ER) to receive tail-        | Yes       |
|   | (Tail-anchored      |         |        |       |                   | protein        | anchored (TA) proteins post-translationally (Vilardi et al. 2011).  |           |
|   | protein insertion   |         |        |       |                   |                |   |           |
|   | receptor WRB)       |         |        |       |                   |                |   |           |
| 2 | 6MZCM               | Q16594  | 0.899  | 0.074 | Homo sapiens      | Transcription  | A significant part of the beginning of RNA polymerase II (Pol II)-  | Yes       |
|   |                     |         |        |       |                   |                | dependent transcription is played by the TFIID basal transcription  |           |
|   | (Transcription      |         |        |       |                   |                | factor complex (Chen et al. 2021)                                   |           |
|   | initiation factor   |         |        |       |                   |                |   |           |
|   | TFIID subunit 9)    |         |        |       |                   |                |   |           |
| 3 | 2NPSC               | Q9JI51  | 0.890  | 0.000 | Rattus norvegicus | Transport      | By interacting with t-SNAREs on the target membrane, the V-         | No        |
|   |                     |         |        |       |                   | protein        | SNARE mediates vesicle transport pathways. It is suggested that     |           |
|   | (Vesicle transport  |         |        |       |                   |                | these interactions mediate certain elements of the specificity of   |           |
|   | through interaction |         |        |       |                   |                | vesicle trafficking and encourage the fusing of the lipid bilayers. |           |
|   | with t-SNAREs       |         |        |       |                   |                | engaged in the movement of vesicles from late endosomes to the      |           |
|   | homolog 1A)         |         |        |       |                   |                | trans-Golgi network. KCNIP1 and KCND2 use an unconventional         |           |
|   |                     |         |        |       |                   |                | RAB1-dependent traffic pathway to the cell surface in conjunction   |           |
|   |                     |         |        |       |                   |                | with VAMP7 (By similarity). Concerned about cytokine output that    |           |

|   |  |        |       |       |   |                      | has risen due to cellular aging (Zwilling et al. 2007).   |     |
|---|--|--------|-------|-------|---|----------------------|---|-----|
| 4 | 6Z6FD  (HDA1 complex subunit 2)                        | Q06629 | 0.888 | 0.111 | Saccharomyces<br>cerevisiae (strain<br>ATCC 204508 /<br>S288c))       | Gene<br>regulation   | Necessary for the HDA1 histone deacetylase complex to function. The lysine residues on the N-terminal portion of the core histones (H2A, H2B, H3 and H4) are deacetylated by the HDA1 histone deacetylase complex. In transcriptional control, cell cycle progression, and developmental processes, histone deacetylation provides a tag for epigenetic repression (Wu <i>et al.</i> 2001). | No  |
| 5 | 5B3DA  (Flagella synthesis protein FlgN)               | P0A1J7 | 0.884 | 0.111 | Salmonella<br>typhimurium (strain<br>LT2 / SGSC1412 /<br>ATCC 700720) | Protein<br>transport | N/A   | No  |
| 6 | 7O3VA (TrwJ protein)                                   | O50331 | 0.884 | 0.185 | Escherichia coli  | Membrane<br>protein  | N/A   | No  |
| 7 | 7T3UA  (Inositol 1,4,5- trisphosphate receptor type 3) | Q14573 | 0.883 | 0.111 | Homo sapiens  | Metal<br>transport   | Inositol 1,4,5-trisphosphate receptor, a second messenger that mediates the release of intracellular calcium (By similarity). Homeostasis of calcium ions within cells is affected (Rönkkö <i>et al.</i> 2020).   | Yes |
| 8 | 7OPCS  (RNA polymerase-associated protein LEO1)        | Q8WVC0 | 0.879 | 0.148 | Homo sapiens  | Transcription        | It is a part of the PAF1 complex (PAF1C), which is involved in the control of embryonic stem cell pluripotency and has a variety of roles during transcription by RNA polymerase II (Kim <i>et al.</i> 2010).   | Yes |
| 9 | 6FKFB (ATP synthase                                    | P00825 | 0.879 | 0.037 | Spinacia oleracea   | Membrane protein     | when a proton gradient is present across the membrane, it converts ADP to ATP. In general, the beta subunits host the catalytic sites   | No  |

|    | subunit beta, chloroplastic)          |        |       |       |                   |           | (Yang et al. 2020).   |    |
|----|---------------------------------------|--------|-------|-------|-------------------|-----------|---|----|
| 10 | 6VQ6I                                 | O6PCU2 | 0.879 | 0.074 | Rattus norvegicus | Proton    | Subunit of the vacuolar(H+)-ATPase (V-ATPase), a multisubunit   | No |
|    | (V-type proton<br>ATPase subunit E 1) |        |       |       | Ü                 | transport | enzyme made up of a membrane-integral complex (V0) that transports protons and a peripheral complex (V1) that hydrolyzes ATP.   |    |
|    |                                       |        |       |       |                   |           | In some cell types, V-ATPase is directed to the plasma membrane, where it is in charge of acidifying the extracellular environment in addition to acidifying and regulating the pH of internal compartments (Abbas <i>et al.</i> 2020). |    |

<sup>\*</sup> TM-score: a metric for assessing the topological similarity of protein structures. \* IDEN: the percentage sequence identity in the structurally aligned region.

epitopes with other protein saliva possibly low. In terms of protein structure, the C3 and D2 epitopes are unique. Since the aim of this research is to produce antibody polyclonal against CYFRA 21-1, the possibility of cross-reaction can be suppressed or abolished using some blocking agent (Lv *et al.* 2020).

#### Proteolytic cleavage analysis

From the proteolytic cleavage analysis using PeptideCutter, there were several enzymes that cleave with both epitope (Shown in Table 7). Data shows the enzymes that causes the most cleavages is Proteinase K, Thermolysin, and Pepsin (pH>2) with 30, 20 and 15 number of cleavages.

The proteolytic cleavage analysis shows that there were several enzymes that may cause cleavage to the epitopes. In general, the cleavage is relatively normal according to the function of each cleavage enzyme. As can be seen, most prominently, the cleavage is carried out by Proteinase K. Proteinase K is known as the most destructive proteinase. Based on this description of the cleavage site, handling the C3 and D2 epitopes when used as antigen must pay attention

to several aspects so that the protease enzymes do not have time to cut the epitope sequence. This can be done by using adjuvants as stabilizers and also paying attention to the buffer used regarding pH and storage temperature (Eladawy *et al.* 2020).

#### CONCLUSION

In this study, we have successfully designed two specific antigen (epitopes) that have the potential to generate antibodies against CYFRA 21-1 *in silico*. The epitopes are C3 and D2 with the sequences QSQLSMKAALEDTLAETEARFGAQLAH and IQALISGIEAQLGDVRADSERQNQEYQ

respectively. Both do not have cross-reactions with other salivary biomarker proteins and have a unique protein structure that has the potential to be used as antigen to produce antibodies for the development of saliva-based immunodiagnostics to diagnose NPC patients. To the best of our knowledge, this is the first reported *in silico* antigen design for salivary biomarkers in detecting NPC. To approve this antigen design, an *in vivo* study needs to be carried out.

Table 7. Enzymes that cleave CYFRA 21-1 sequence

| Name of enzyme                       | No. of cleavages | Positions of cleavage sites  |
|--------------------------------------|------------------|--|
| Arg-C proteinase                     | 4                | 20 43 48 55  |
| Asp-N endopeptidase                  | 3                | 11 40 44   |
| Asp-N endopeptidase + N-terminal Glu | 9                | 10 11 15 17 35 40 44 46 51   |
| CNBr                                 | 2                | 6 57   |
| Chymotrypsin-high specificity        | 2                | 21 53  |
| Chymotrypsin-low specificity         | 12               | 4 6 10 14 21 25 27 31 39 53 56 57  |
| Clostripain                          | 4                | 20 43 48 55  |
| Formic acid                          | 3                | 12 41 45   |
| Glutamyl endopeptidase               | 6                | 11 16 18 36 47 52  |
| LysC                                 | 1                | 7  |
| LysN                                 | 1                | 6  |
| Pepsin (pH 1,3)                      | 13               | 3 4 10 13 14 21 24 25 30 31 38 39 56   |
| Pepsin (pH >2)                       | 15               | 3 4 10 13 14 21 24 25 30 31 38 39 52 53 56   |
| Proteinase K                         | 30               | 4 8 9 10 11 13 14 15 16 17 18 19 21 23 25 26 28 30 31 32 35 36 37 39 42 44 47 52 53 56 |
| Staphylococcal peptidase I           | 6                | 11 16 18 36 47 52  |
| Thermolysin                          | 20               | 3 5 7 8 9 13 14 20 22 24 25 27 29 30 31 34 38 43 55 56                                 |
| Trypsin                              | 5                | 7 20 43 48 55  |

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#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

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#### **Supplementary-1**

CLUSTAL 2.1 multiple sequence alignment -----TOA D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P16145 | IL8\_HUMAN sp | P60174 | TPIS\_HUMAN sp | P0DTE8 | AMY1C\_HUMAN -----MHLLGFFSVACSLLAAALLPGPREA -----MAEVPELASE MKLFWLLFTIGFCWAQYSSNTQQGRTSIVHLFEWRWVDIALECERYLAPK QLS-MKAALEDTLAETEARFGAQLAH-----LTSGIEAQLGDVRADSERQNQEYQ--PAAAAAFESGLDLSDAEPDAGEATAYASKOLEEQLRSVSSVDELMTVLYP
MAAYYSGNEDDLFFEADGPKQMKCSFQDLDLCPLDGGTQLRISDHTVJSKG
LLAAFLISAALCEGAVLPRSAKELRCQCTKTVSKPFHPKFIKELRVIESG D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P10145 | IL8\_HUMAN sp P60174 TPIS\_HUMAN GGNNKMNGRKQSLGELIGTLNAAKVPADTEVVCAPPTAYIDFARQKLDPK GFGGVQVSPPNENVAIHNPFRPWHERYQPVSYKLCTRSGNEDEFRNMVTR sp PODTES AMY1C\_HUMAN D2 ..... D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | TL1B\_HUMAN sp | P10145 | TL8\_HUMAN sp | P60174 | TPTS\_HUMAN sp | P0DTE8 | AMY1C\_HUMAN EYWKMYKCOLRKGGWOHNREOANLNSRTEETIKFAAAHYNTEILKSIDNE FRQAASVVVAMDKLRKMLVPCPQTFQENDLSTFFPFIFEEPIFFDTWDN
PHCANTEIIVKLSDGRELCLDPKENWVQRVVEKFLKRAENS-----IAVAAONCYKVTNGAFTGEISPGMIKDCGATWVVLGHSERRHVFGESDEL CNNVGVRIYVDAVINHMCGNAVSAGTSSTCGSYFNPGSRDFPAVPYSGWD WRKTQCMPREVCIDVGKEFGVATNTFFKPPCVSVY---RCGGCCNSEGLQ sp P49767 VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P10145 | IL8\_HUMAN sp | P60174 | TPIS\_HUMAN EAYVHDAPVRSLNCTLRDSQQKSLVMSGPYELKAL---HLQGQDMEQQVV IGQKVAHALAEGLGVIACIGEKLDEREAGITEKVV---FEQTKVIADN-sp PODTES AMY1C\_HUMAN FNDGKCKTGSGDIENYNDATQVRDCRLSGLLDLALGKDYVRSKIAEYMNH C3 -----D2 sp|P49767|VEGFC\_HUMAN sp|P01584|IL1B\_HUMAN CMNTSTSYLSKTLFEITVPLSQGPKPVTISFANHTSCRCMSKLDVYRQVH FSMS-----FVQGEESNDKIPVALGLKEKNLYLSCVLKDDKPT sp P61384 ILIB\_HUMAN sp P60174 TPIS\_HUMAN sp P60TE8 AMY1C\_HUMAN -----VKDWSKVVLAYEPVWAIGTGKTATPQQAQEVH LIDIGVAGFRIDASKHMWPGDIKAILDKLHNLNSNWFPEGSKPFIYQEVI D2 D2 Sp | P49767 | VEGFC\_HUMAN Sp | P01584 | TL1B\_HUMAN Sp | P10145 | TL8\_HUMAN Sp | P60174 | TPIS\_HUMAN Sp | P0DTE8 | AMY1C\_HUMAN SIIRRSLPATLPQCQAANKTCPTNYMWNNHICRCLAQEDFMFSSDAGDDS LQLESVDPKNYPKKKMEKRFVFNKIEINNKLEFESAQFPNWYISTSQAEN EKLRGWLKSNVSDAVAOSTRIIYGGSVTGATCKELASOPDVDGFLVGGAS DLGGEPIKSSDYFGNGRVTEFKYGAKLGTVIRKWNGEKMSYLKNWGEGWG D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P10145 | IL8\_HUMAN sp | P60174 | TPIS\_HUMAN TDGFHDICGPNKELDEETCOCVCRAGLRPASCGPHKELDRNSCOCVCKNK MPVFLGGTKGGQDITDFTMQFVSS-----LKPEFVDIINAKO----sp PODTES AMY1C\_HUMAN FMPSDRALVFVDNHDNQRGHGAGGASILTFWDARLYKMAVGFMLAHPYGF ----sp P49767 VEGFC\_HUMAN LFPSQCGANREFDENTCQCVCKRTCPRNQPLNPGKCACECTESPQKCLLK SP | P01584 | IL1B\_HUMAN SP | P10145 | IL8\_HUMAN SP | P60174 | TPIS\_HUMAN SP | P0DTE8 | AMY1C\_HUMAN TRVMSSYRWPRYFENGKDVNDWVGPPNDNGVTKEVTINPDTTCGNDWVCE D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P10145 | IL8\_HUMAN sp | P60174 | TPIS\_HUMAN sp | P0DTE8 | AMY1C\_HUMAN GKKFHHQTCSCYRRPCTNRQKACEPGFSYSEEVCRCVPSYWKRPQMS---..... HRWROIRNMVNFRNVVDGOPFTNWYDNGSNOVAFGRGNRGFIVFNNDDWT C3 D2 52 Sp | P49767 | VEGFC\_HUMAN 5p | P91584 | IL1B\_HUMAN 5p | P19145 | IL8\_HUMAN 5p | P69174 | TPIS\_HUMAN 5p | P60TE8 | AMY1C\_HUMAN -----FSLTLQTGLPAGTYCDVISGDKINGNCTGIKIYVSDDGKAHFSISNSAED D2 sp | P49767 | VEGFC\_HUMAN sp | P01584 | IL1B\_HUMAN sp | P16145 | IL8\_HUMAN sp | P60174 | TPIS\_HUMAN sp | P0DTE8 | AMY1C | HUMAN ----------PFIAIHAESKL

Figure 1. Sequence alignment with other protein saliva.

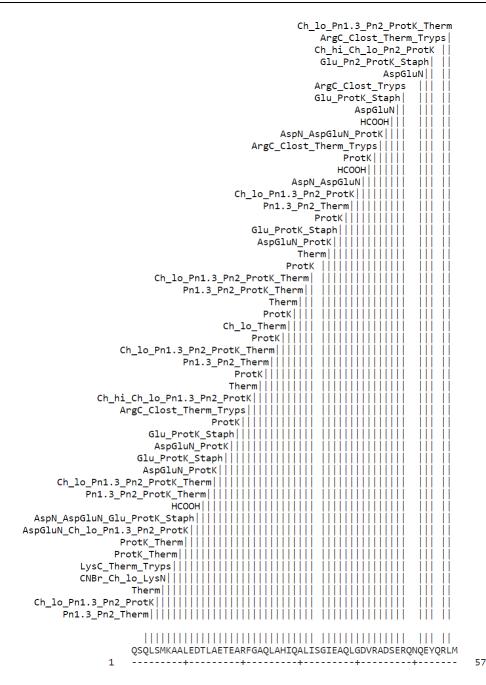


Figure 2. Proteolytic cleavages in CYFRA 21-1.