

# Antimicrobial potency of toothpaste containing gambir (*Uncaria gambir*) extract

Siti Rusdiana Puspa Dewi<sup>1\*</sup>  
 Pudji Handayani<sup>2</sup>  
 Danica Anastasia<sup>3</sup>  
 Shania Tri Maulina<sup>4</sup>

<sup>1</sup>Department of Biomedical Science,  
 Dentistry Study Programme,  
 Medical Faculty, Universitas  
 Sriwijaya, Palembang, Indonesia

<sup>2</sup>Department of Oral Medicine,  
 Dentistry Study Programme,  
 Medical Faculty, Universitas  
 Sriwijaya, Palembang, Indonesia

<sup>3</sup>Department of Operative  
 Dentistry, Dentistry Study  
 Programme, Medical Faculty,  
 Universitas Sriwijaya, Palembang,  
 Indonesia

<sup>4</sup>Student of Dentistry Study  
 Programme, Medical Faculty,  
 Universitas Sriwijaya, Palembang,  
 Indonesia

\*Correspondence:  
[sitirusdiana@fkk.unsri.ac.id](mailto:sitirusdiana@fkk.unsri.ac.id)

Received: 29 May 2023  
 Revised: 06 July 2023  
 Accepted: 31 July 2023  
 Published: 31 July 2023  
 DOI: [10.24198/pjd.vol35no2.47130](https://doi.org/10.24198/pjd.vol35no2.47130)

p-ISSN 1979-0201  
 e-ISSN 2549-6212

Citation:  
 Dewi SRP, Handayani P, Anastasia  
 D, Maulina ST. Antimicrobial  
 potency of toothpaste containing  
 gambir (*Uncaria gambir*) extract.  
 Padj J Dent, July. 2023; 35(2): 98-  
 105.

## ABSTRACT

**Introduction:** The biggest problem in oral health is dental caries and periodontal disease. The way to prevent dental and mouth problems is to keep oral hygiene. One of them is adequate plaque control. Plaque control can be done by brushing teeth and using toothpaste. Currently, many herbal ingredients have been used as antibacterials such as gambir (*Uncaria gambir*). Gambir mainly contains catechins and tannins, which have the ability to inhibit the growth of bacteria. The purpose of this study was to determine the antimicrobial potential of toothpaste containing gambir (*Uncaria gambir*). **Methods:** The total sample was 30, and it divided into three groups. Samples were toothpaste containing ten percent of *Uncaria gambir* extract, fluoride, placebo toothpaste, *Streptococcus mutans* (Gram-positive), *Porphyromonas gingivalis* (Gram-negative), and *Candida albicans*. Antimicrobial activity of toothpaste with gambir (*Uncaria gambir*) extract against microorganisms was observed with the formation of inhibitory zone diameter in agar. Data were analyzed by using SPSS with one-way ANOVA and Tukey test. **Results:** The results showed that toothpaste containing gambir extract could inhibit the *Streptococcus mutans* ( $p < 0.05$ ), *Porphyromonas gingivalis* ( $p < 0.05$ ), *Candida albicans* ( $p < 0.05$ ) compared significantly with placebo ( $p < 0.05$ ). Antimicrobial toothpaste containing gambir extract had no different effect from toothpaste containing fluoride. However, toothpaste containing gambir extract had significantly different effect than toothpaste containing fluoride. **Conclusion:** It can be concluded that toothpaste containing gambir (*Uncaria gambir*) extract has antimicrobial potential against *Streptococcus mutans*, *Porphyromonas gingivalis*, and *Candida albicans*. These findings showed that the extract of *Uncaria gambir* possesses potent antimicrobial activity.

## KEYWORDS

antimicrobial, *candida albicans*, *porphyromonas gingivalis*, *streptococcus mutans*, *uncaria gambir*

## INTRODUCTION

Dental caries, Periodontal disease, and oral mucosal lesions are the main problems worldwide.<sup>1</sup> Those diseases are caused by microbes that live in the oral cavity. Dental caries is a chronic disease caused by complex interactions among acid-producing bacteria, tooth surfaces, fermentable carbohydrates, and time.<sup>2</sup> *Streptococcus mutans* is a bacterium that is mostly found in the dental cavity.<sup>3</sup> *Streptococcus mutans* is a Gram-positive, cocci-shaped, and facultative anaerobic bacterium. This type of bacteria can survive in an acidic environment, produce glycosyltransferases (GTFs), and form glucan. Glucan plays a role in the attachment and increases bacteria colonies, then causes dental plaque.<sup>4</sup>

The most common bacteria involved in the pathogenesis of periodontal disease is *Porphyromonas gingivalis*.<sup>5,6</sup> *Porphyromonas gingivalis* is an anaerobic Gram-negative bacterium that plays an important role in the third phase of plaque formation of secondary colonization and microbial maturation. These bacteria produce virulence factors such as proteases that can destroy immunoglobulins, complement factors, heme-sequestering, hemolysin, and collagenase. Other virulence factors are fimbriae (FimA), gingipain, and lipopolysaccharide (LPS).<sup>7</sup>

Gingipain and lipopolysaccharide (LPS) are virulence factors of *Porphyromonas gingivalis* used to invade the periodontium tissue. Fimbriae A (FimA) helps *Porphyromonas gingivalis* adhere to subgingival bacteria such as *Streptococcus* and *Actinomyces* species.<sup>6,7</sup> Various surface components of *Porphyromonas gingivalis* enable it to interact with host cells, such as neutrophils, epithelial cells, fibroblasts, etc., thus making it easier for bacteria to grow, acquire nutrients, colonize, and form plaque biofilms.<sup>7</sup> Their productions influence the migration of PMN leukocytes, weaken the immune system, and lead to the damage of periodontal tissue.<sup>6,7</sup>

The prevalence of oral mucosal lesions in the global population is approximately 64.7%.<sup>8</sup> Lesions of the oral mucosa are characterized by the alteration of color, surface, shape, mucosal integrity, and swelling. Microbial infections, such as candida, mostly contribute to these lesions.<sup>8</sup> *Candida albicans* is an opportunistic microbe that is found in oral candidiasis. This lesion can be found in every oral mucosa, but the most common locations are buccal, buccal folds, oropharynx, and tongue.<sup>9</sup> The pathogenicity of this fungus as a human pathogen is determined through several mechanisms, such as biofilm formation, mediating adhesion and invasion of human cells, secreting the hydrolyses, yeast-to-hypha transition, contact sensing, thigmotropism, and phenotypic switching.<sup>10</sup>

To prevent oral and dental problems, people should improve oral hygiene by reducing the accumulation of plaque, called plaque control.<sup>2</sup> The aim of plaque control is to eliminate bacteria in dental plaque.<sup>2,3</sup> Plaque control is classified into mechanical plaque control and chemical plaque control. Mechanical plaque control can be done by brushing teeth, using dental floss, and interdental brush, while chemical plaque control can be achieved by using toothpaste and mouthwash.<sup>11</sup> Optimal plaque control can be achieved by the combination of mechanical and chemical plaque control. Brushing teeth with toothpaste is the first step to controlling oral hygiene.<sup>2,11</sup>

The functions of toothpaste are to eliminate microbes that live in the oral cavity, control the plaque, polish the tooth surface, strengthen the teeth, prevent bad breathing, and maintain the hardness of soft tissue in the oral cavity.<sup>12</sup> Toothpaste is a thick-mass mixture of powder and liquid. Toothpaste contains active and inactive ingredients.<sup>13</sup> The active ingredients of toothpaste provide a therapeutic effect, and the inactive ingredients thicken toothpaste formulations, bind to toothpaste components, and have a certain color or taste. Among the inactive ingredients include abrasive agents, humectants, binding agents, surfactants, preservatives, sweeteners, fragrances, and solvents.<sup>14</sup> The active ingredients used are antibacterial agents, such as fluoride or herbs.<sup>15</sup> Several studies have proven that herbal ingredients have benefits for reducing the number of bacteria and fungi that cause oral disease.<sup>16</sup> Nuraskin et al. showed that herbal toothpaste was more effective compared to conventional toothpaste.<sup>17</sup> The American Dental Association (ADA) and the Indonesian National Standard (SNI) recommend the use of herbal ingredients because herbal ingredients are relatively safe, cause no side effects, and are cheap and easy to find.<sup>18</sup> One of herbal ingredients that can be used to improve the function of toothpaste and have high biocompatibility is Gambir (*Uncaria gambir*).<sup>19</sup>

Gambir (*Uncaria gambir*) is a tropical plant that is mostly found in Indonesia. The biochemical compounds in Gambir extract are catechins (7-33%), catechin tannic acid (20-55%), pyrocatechol (20-30%), fluorescent Gambir (1-3%), red catechu (3-5%), quercetin (2-4%), certain oils (1-2%), wax (1-2%), and small concentrations of alkaloids.<sup>20</sup> Catechins, the main contents of Gambir, can damage the cell membranes or cell walls by interfering with the cell permeability of bacteria.<sup>21</sup> Melia et al. (2015) reported that *Uncaria gambir* extract has antimicrobial activity against some Gram-positive bacteria (*Staphylococcus aureus*) and Gram-negative (*E. coli* and *Salmonella* sp).<sup>22</sup> Catechins are antioxidants that have a mechanism of action in the form of decreasing the mechanism of ergosterol synthesis in *Candida albicans* cell walls which can further suppress the growth of *Candida albicans*.<sup>23</sup> Ifora<sup>24</sup> stated that ethanol extract from gambir leaves (*Uncaria gambir* Roxb) could inhibit the growth of *Candida albicans*. Another study (2016) also reported that *Uncaria gambir* extract was capable of reducing the decline of micropores on the enamel surface, minimizing the decline of calcium weight, and reducing bacteria colonies.<sup>25</sup> It was also stated that best conditions of gambir extraction process was by using a solvent ethyl acetate<sup>20</sup>, the addition of gambir extract to toothpaste can be used in preventing dental caries. Besides that, the addition of gambir toothpaste can also be used as an alternative to toothpaste for users who are sensitive to fluoride. The purpose of this study was to know the antimicrobial potential of toothpaste containing gambir extract (*Uncaria gambir*).

## METHODS

The study was *in vitro* experimental laboratory with a post-test-only control group design. It was conducted at Balai Besar Laboratorium Kesehatan (BBLK) of South Sumatra Province. *Streptococcus mutans* ATCC 25175, *Porphyromonas gingivalis* ATCC 33277, and *Candida albicans* ATCC 10231 obtained from BBLK Palembang. The object of this study was toothpaste containing gambir 10%. Isromarina et al stated that ethyl acetate *Uncaria gambir* extracts at 10% concentration could inhibit the growth of *Vibrio cholera*.<sup>26</sup> Dewi et al.<sup>27</sup> Reported that 10% *Uncaria gambir* extract has an antiseptic effect on gingival wounds in rats. Gambir leaves were collected from Babat Toman, Sekayu District of South Sumatera, and had been identified and authenticated by Faculty of Agriculture, Universitas Sriwijaya, Indonesia.

The samples size was 90, it was divided into 3 groups. Each group consisted of 3 subgroups. The sample size of the subgroup was 10. Group I was Gram-positive bacteria (*Streptococcus mutans*), Group II was Gram-negative bacteria (*Porphyromonas gingivalis*), and Group III was *Candida albicans*. Those 3 groups were divided into 3 subgroups; subgroup 1 was toothpaste without active ingredients, or a placebo as negative control; subgroup 2 was toothpaste containing fluoride as positive control; and subgroup III was toothpaste containing Gambir extract (*Uncaria gambir*).

The preparation of Gambir extract started when Gambir leaves were dried in the sun for 48 hours without direct exposure to sunlight. 60 grams of dried Gambir was mashed with a size of 40-60 mesh. The mashed Gambir was then wrapped in filter paper and put into the soxhlet tube. 98% of ethyl acetate, 300 ml, was put into the soxhlet tube, and heated at 77 C for 7 hours. The solution

obtained was evaporated using a rotary evaporator for 2 days. The process of evaporation produces 100% of Gambir extract. The extract was diluted with distilled water to produce 10% of the extract.<sup>20</sup>

The preparation of toothpaste was accomplished by heating a mortar with hot water, and drying it. Sodium Carboxymethylcellulose (CMC) was dissolved in hot water (p to u20 times the weight of sodium CMC) and left in the mortar for 15 minutes while stirring until homogeneous (mass A). Calcium carbonate and sodium benzoate were crushed, and glycerin was added, and then mixture was stirred until homogeneous (mass B). Mass B was added to the mortar containing mass A, and they were thoroughly combined. In this part, natrium fluoride was added for Group II and Gambir extract for Group III, and they were stirred until homogeneous. Saccharin was added to the pasta mass and stirred until homogeneous. Sodium lauryl sulfate was added to the paste mass. The toothpaste mixture was blended and then distributed evenly to be homogenous. The toothpaste homogeneity test was carried out by smearing the toothpaste on a glass object before putting it in the ointment pot.<sup>28</sup>

To prepare *Streptococcus mutans* in agar media, 40 grams of blood agar base were dissolved in 1000 ml of distilled water and then heated and stirred until homogeneous. The agar media was then sterilized using an autoclave at 121°C for 15 minutes. 5% of the total base medium (blood agar base) of Sheep blood was added to the agar medium at 45-60°C and then shaken until homogeneous. 10-15 ml of blood medium was poured into a petri dish and left to become solid.<sup>3</sup>

Isolates of *Streptococcus mutans* were taken as 1-2 oese and suspended in a 0.9% sodium chloride (NaCl) solution in a sterile test tube to meet Mc Farland standard of 0.5 (10-8 CFU/ml). To avoid excess liquid, a sterile cotton swab was dipped into the suspension and then squeezed into a test tube. In a petri dish, the sterile cotton swab was smeared evenly on the blood agar medium.<sup>3</sup>

The preparation of Phorphyromonas media began by placing the brucella media to dissolve and homogenize. The media was sterilized at 121°C with a pressure of 2 atm for 15 minutes using an autoclave. The sterile media was cooled at a temperature of 40-45°C, then mixed with 5% sheep blood and homogenized. The media was transferred to a petri dish.<sup>29</sup>

Isolates of *Porphyromonas gingivalis* were taken and put in a test tube. 2 ml of 0.45% NaCl was added to the test tube. Next, the test tube was put in an anaerobic jar and then incubated for 48 hours at 37°C using an incubator. After that, the dilution was carried out by adding sterile distilled water and homogenizing it using a vortex. Bacterial turbidity was equivalent to 0.5 McFarland or 1.5 X 10<sup>8</sup> CFU/ml.<sup>30</sup> 65 gr of the medium was suspended in a liter of distilled water to prepare the *Candida albicans* medium. The medium was heated with frequent agitation and boiled for one minute to completely dissolve it, then put into an autoclave at 121°C for 15 minutes. It was cooled to 45 to 50°C and poured into Petri dishes or tubes for slanting.<sup>31</sup>

The culture of *Candida albicans* on Sabouraud Dextrose Agar medium was incubated at room temperature for 2 x 24 hours (or two 24-hour cycles). The cultures were then extracted with an ose needle and suspended in a test tube filled with 0.9% NaCl. The fungal suspension in the test tube was homogenized, and the turbidity was adjusted using a densitometer with a MacFarland standard of 0.5 (1-5 x 10<sup>6</sup> CFU/ml).<sup>31</sup>

1.5 gr of toothpaste was diluted with 5 ml of distilled water using a sterile spoon. The toothpaste solution was put in a Petri dish. The paper disc was taken using tweezers and dipped in a Petri dish filled with toothpaste solution for 30 minutes. The disc paper is placed in a petri dish containing each medium and microbe according to the treatment group. The Petri dish that had been treated with disc paper was incubated for 24 hours at 37°C for bacteria and 48 hours at 37°C for candida. The Petri dish was removed from the incubator, then the inhibition zone was measured using a calliper.<sup>32</sup>

Data analysis was performed with the SPSS program. The obtained data were analyzed by normality and homogeneity tests using the Shapiro-Wilk test and Levene's test. The parametric test was carried out using the One-Way ANOVA statistical test to determine whether there was an effect of Gambir toothpaste (*Uncaria gambir* Roxb.) on the Gram-positive, Gram-negative, and Candida bacteria. The analysis was continued with the post-hoc Tukey test to determine the significance. The results of the study were stated to be statistically significant if the p-value <0.05.

## RESULTS

The inhibitory zones of each sample were evaluated. All the groups and subgroups showed inhibitory zones, except placebo toothpaste.

**Table 1.** The average diameter of zone inhibition against microbes

Groups	Subgroups	n	Zone of inhibition (mm)
<i>Streptococcus mutans</i>	Placebo toothpaste	10	0.001 ± 0.01
	Toothpaste containing fluoride	10	9.58 ± 0.52
	Toothpaste containing Gambir	10	9.13 ± 0.62
<i>Porphyromonas gingivalis</i>	Placebo toothpaste	10	0.001 ± 0.01
	Toothpaste containing fluoride	10	14.22 ± 0.42
	Toothpaste containing Gambir	10	7.65 ± 0.12
<i>Candida albicans</i>	Placebo toothpaste	10	0.001 ± 0.01
	Toothpaste containing fluoride	10	1.37 ± 0.66
	Toothpaste containing Gambir	10	1.82 ± 0.48

Table 1 described the average diameter of inhibitory zone of toothpaste against Gram-positive (*Streptococcus mutans*), Gram-negative (*Porphyromonas gingivalis*), and *Candida albicans*. According to the results, significant antimicrobial activities of toothpaste with fluoride and toothpaste with Gambir extract were shown against these bacteria and *Candida*. were shown on toothpaste containing fluoride-containing toothpaste showed the highest zones of inhibition of *Streptococcus mutans* and *Porphyromonas gingivalis*, while toothpaste with Gambir extract came in second. While the highest zone of inhibition of *Candida albicans* was given by toothpaste containing Gambir, followed by toothpaste containing fluoride. Placebo did not show any inhibitory zones in all subgroups.

The normality test using Shapiro Wilk and the homogeneity test using Levene's test showed p-values >0.05. It meant that all data were normally distributed and homogenous (identical). Data analysis was continued using the One-way ANOVA test. According to Table 2, the toothpaste treatment in all groups obtained p<0.05, which meant that there was a significant difference in the inhibitory zone among the treatment subgroups. The analysis was continued with the post-hoc Tukey test to find out which of the four treatment groups had statistically significant differences (Tables 3,4, and 5).

**Table 2.** One-way ANOVA test results

Groups	p
<i>Streptococcus mutans</i>	0.001*
<i>Porphyromonas gingivalis</i>	0.001*
<i>Candida albicans</i>	0.001*

\*significant (p<0.05)

Table 3 showed that there was a significant difference between placebo and toothpaste containing fluoride; and placebo with toothpaste containing Gambir. However, there was no significant difference between toothpaste containing fluoride and toothpaste containing Gambir. It can be concluded that the potency of the antibacterial agent of toothpaste with Gambir extract had the same ability as one with fluoride against Gram-positive bacteria (*Streptococcus mutans*).

**Table 3.** Post-Hoc Tukey test from the toothpaste against *Streptococcus mutans*

Sub groups	Placebo	Toothpaste containing fluoride	Toothpaste containing gambir
Placebo		0.001*	0.001*
Toothpaste with fluoride	0.001*		0.68
Toothpaste with Gambir	0.001*	0.68	

\*significant (p<0.05)

Table 4 showed that the comparison of all subgroups had significant differences (p<0.05). It described that toothpaste containing gambir had a potential effect as an antibacterial agent against Gram-negative (*Porphyromonas gingivalis*) but not as strong as toothpaste containing fluoride.

**Table 4.** Post-Hoc Tukey test from the toothpaste against *Porphyromonas gingivalis*

Sub groups	Placebo	Toothpaste containing fluoride	Toothpaste containing gambir
Placebo		0.001*	0.001*
Toothpaste containing fluoride	0.001*		0.001*
Toothpaste containing Gambir	0.001*	0.001*	

\*significant (p<0.05)

Table 5 showed that there was significant difference among all subgroups. In contrast, there was no significant difference between toothpaste containing fluoride and toothpaste containing Gambir. Therefore, the antibacterial effect of Gambir-containing toothpaste had the same ability as fluoride-containing toothpaste against *Candida albicans*.

**Table 5.** Post-Hoc Tukey test from the toothpaste against *Candida albicans*

Sub groups	Placebo	Toothpaste containing fluoride	Toothpaste containing gambir
Placebo		0.001*	0.001*
Toothpaste containing fluoride	0.001*		1.00
Toothpaste containing gambir	0.001*	1.00	

\*significant (p<0.05)

## DISCUSSION

According to Davis and Stout (1971), the diameter of the inhibition zone is determined by the antimicrobial activity of active substances. Substances with an inhibition zone diameter of less than 5 mm were categorized as weak antimicrobials. Substances with inhibition zone diameters of 5-10 mm were categorized as having moderate antimicrobial properties. And substances with an inhibition zone diameter of 11-20 mm were categorized as strong antimicrobial agents.

The study indicated that toothpaste containing Gambir was categorized as having moderate antibacterial properties against *Streptococcus mutans* and *Porphyromonas gingivalis*, and was categorized as having weak antimicrobial properties against *Candida albicans*. Toothpaste containing fluoride was categorized as having moderate antibacterial properties against *Streptococcus mutans*, strong antibacterial agents against *Porphyromonas gingivalis*, and weak antimicrobial properties against *Candida albicans*.

The results from Table 1 showed that Gambir-containing toothpaste increased the antimicrobial ability against *Streptococcus mutans*, *Porphyromonas gingivalis*, and *Candida albicans*. The main components of Gambir extracts were catechins and tannins.<sup>33</sup> Inmawaty et al. stated that the extract of Gambir containing catechins had an inhibition zone against *Streptococcus mutans*.<sup>34</sup> Catechins are phenolic group compounds that play important role in denaturing and precipitating bacterial cell proteins, and inactivating *Streptococci* enzymes.<sup>35</sup> Catechins interact with peptidoglycan and change the structure of the bacterial cell wall.<sup>36</sup> This process increases the permeability of the bacterial cell membrane, which is followed by cell membrane damage, and then the bacteria will lyse and die.<sup>35</sup> The different results of antimicrobial activities among different microbes (Table 3, 4 and 5) were due to the distinction of cell wall and its composition. There was a difference between Gram-positive and Gram-negative bacterial cell walls. The composition of the peptidoglycan layer of Gram-positive bacteria has a thicker peptidoglycan layer than that of Gram-negative.<sup>37</sup> Therefore, the toothpaste containing Gambir extract is more effective against *Streptococcus mutans* than against *Porphyromonas gingivalis*, as described in this study. Taylor stated that catechins tend to have less activity against Gram-negative than they are against Gram-positive bacteria.<sup>38</sup>

Catechins are able to work competitively with glucosyltransferase (GTF) enzymes, preventing the bacteria from synthesizing extracellular polysaccharides (glucans) necessary for *Streptococcus mutans* attachment and colonization.<sup>39</sup> Catechins also play a role in inhibiting the metabolism of *Streptococcus mutans* at the glycolysis stage.<sup>35</sup> Catechins bind to the catalyst to inhibit the activity of the enzyme lactate dehydrogenase.<sup>36</sup> This activity makes it difficult for bacteria convert pyruvate to lactic acid.<sup>35,36</sup> Xu et al. reported that catechins inhibited not only lactate dehydrogenase activity but also F1Fo-ATPase.<sup>40</sup> F1Fo-ATPase activity plays an important role in the virulence factor of *Streptococcus mutans*.<sup>37</sup> This role is carried out by pumping protons from the cytoplasm to the bacterial cell membrane to maintain pH homeostasis and preserve the bacteria's enzymatic function.<sup>37</sup> Catechins in gambier extract inhibit the activity of the F1Fo-ATPase enzyme, resulting in an increase in cytoplasmic acidity, which in turn interferes with bacteria's cell membrane.<sup>39</sup>

Catechins inhibit the expression of some genes involved in host colonization, such as fimbriae A (*fimA*), tissue destruction, and heme acquisition (*hem*).<sup>6</sup> Reducing the expression of *fimA* may also help reduce inflammation because this virulence factor has the ability to induce host cells to produce cytokines.<sup>7</sup> Catechins also increased the expression of the stress protein *htrA* gene. This gene is known to be responsible for resisting oxidative stress in *Porphyromonas gingivalis*, helping them survive under stressful conditions and causing stress on the bacteria.<sup>40</sup>

Catechins penetrate deeper into the lipid bilayer interface by absorption. It suggests that catechins regulate membrane partitioning by hydrogen bonding between the catechins and membrane lipids, specifically between the hydroxyl groups of the catechin and the oxygen atom of the lipids.<sup>36</sup> Catechins not only interfere with the cell membrane and cell wall of *Porphyromonas gingivalis*, but also affects the growth and adhesion of *Porphyromonas gingivalis*, interrupt the biofilm formation, weaken its invasiveness to host cells and tissues, and reduce the production of volatile sulfur compounds and causing oral halitosis by inhibition of relevant virulence factors such as fimbriae, collagenase, gingival protease, toxic end metabolites.<sup>36,41</sup> Lestari reported that Gambir catechins affected the amount of *Actinobacillus actinomycetemcomitans* (AA) in periodontitis mice.<sup>42</sup> Additionally, Sari and Deynalisa also reviewed that gargling with 1 gram gambir-boiling water was effective in treating gingivitis.<sup>43</sup>

Other active substances found in Gambir extract (*Uncaria gambir* Roxb.) are tannins.<sup>26</sup> Tannins have an antibacterial action due to their ability to deactivate bacterial adhesins, inhibit both the action of enzymes and protein transport in the cell envelope.<sup>44</sup> The mechanism of action of tannins as an antibacterial agent, among others, is through the destruction of bacterial cell membranes due to the toxicity of tannins and the formation of metal ion complex bonds from tannins, which play a role in tannin toxicity.<sup>44</sup> Tannins also have the ability to neutralize toxins in bacteria.<sup>45</sup>

Dewi et al, Reported that tannins play a role in inhibiting bacterial cell metabolism by inducing ionic bonds and causing toxicity, thereby disrupting cell membrane permeability and denaturing proteins.<sup>25,27</sup> Eolia reported that tannins isolated from *Ficus carica* Linn. had the ability to destroy *Porphyromonas gingivalis*.<sup>45</sup> Ho et al, Revealed that tannin compounds were proven to be able to inhibit the growth of bacteria that cause periodontitis, one of which is *Porphyromonas gingivalis*.<sup>46</sup> Choi et al. stated that tannins could reduce the cytotoxicity of bacterial toxins so that bacterial pathogenesis decreased.<sup>47</sup>

This study also presented that toothpaste containing gambir had the ability to inhibit *Candida albicans* and statistically had no different significant effect on toothpaste containing fluoride. The antimycotic activity of gambir is carried out through several underlying mechanisms, including plasma membrane disruption, the induction of mitochondrial dysfunction, inhibiting cell wall formation and division, interfering with RNA and protein synthesis, and affecting the efflux-mediated pumping system.<sup>48</sup> Catechins, an active metabolite from gambir, are known as antioxidants, which are capable of inhibiting the growth of *Candida albicans* by reducing the mechanism of ergosterol synthesis.<sup>24</sup> The mechanism of action of phenol is based on denaturation and precipitating microbial cell proteins, so the antifungal effect of gambir is effective against *Candida albicans*.<sup>25</sup> Furthermore, excess production of reactive oxygen species (ROS) induces severe oxidative stress on the cell, which increases the



permeability of membrane cells, injures nucleic acids, and leads to the oxidation of fatty acids and amino acids.<sup>48</sup> ROS produces lipid peroxidation, resulting in disruption of lipid bilayer, altering membrane potentials, progressing membrane permeability, and interfering with phospholipids.<sup>49</sup> Catechins and tannins found in gambir can change cell wall formation by inhibiting the synthesis of glucans and chitin.<sup>48</sup>

Catechins inhibit *Candida albicans* nucleic acid synthesis. Study analysis by flow cytometry showed that catechins exhibited the inhibition of FCS-induced hyphal formation.<sup>48</sup> Catechins enter the cell through active transport. After they reach the nucleus, they disrupt DNA, RNA, and protein synthesis.<sup>49</sup> Tannins have the ability to inhibit the mitochondrial electron transport chain and diminish the potential of cell membranes. Tannins obstruct the proton pumps and reduce ATP synthesis. This condition causes cell death.<sup>50</sup> Rahayuningsih et al, stated that gambir was effective as antifungal agents.<sup>51</sup> Another study also reported that polyphenolic compounds found in *Uncaria gambir* had a potential effect as antibacterial and antifungal agents.<sup>24</sup>

Toothpaste containing fluoride as positive control had been proven as an antimicrobial agent to maintain oral hygiene. Fluoride inhibits glycolytic enzyme enolase from acidic bacteria and heme-based peroxidase enzymes by binding fluoride to heme.<sup>52</sup> Fluoride involves the formation of metal-fluoride complexes, such as  $AlF_4^-$ .<sup>53</sup> These complexes are responsible for fluoride inhibition of proton-translocating F-ATPases and are thought to act by mimicking phosphate to form complexes with ADP at the reaction centers of the enzymes.<sup>53</sup> Another mode of action is to enhance membrane permeabilities to protons and disrupts the functioning of F-ATPases in exporting protons. Fluoride reduces the acid tolerance of oral bacteria, especially *Streptococcus mutans*.<sup>52</sup> In Gram-negative, fluoride inhibits heme catalase, penetrates cells, and causes cell death.<sup>53</sup> A good penetration of bacteria cell walls will ease the active compounds to kill the cell. Fluorides are compounds that combine with some positively charged counterparts.<sup>54</sup> The anion porphyrins of Gram-negative bacteria will bind with the positive charged compounds, and disorganize the outer membrane structure, so the fluor ion killing effect can take place.<sup>53,54</sup>

Toothpaste containing fluoride has long been known as an antifungal agent. Fluoride blocks the functions of key metabolic enzymes and leads to the growth of candida. The mode of action is by interfering with cell membrane integrity, causing leakage of cytoplasmic constituents so that cell growth will be interrupted.<sup>55</sup> Mozayani et al. Reported that chlorhexidine had significantly higher anticandidal activity against *Candida albicans* compared to fluoride.<sup>56</sup> Adwan et al. stated that parodontax (toothpaste containing herbal ingredients) showed a significant difference ( $P < 0.001$ ) against *Candida albicans* isolates compared to toothpaste containing sodium fluoride.<sup>57</sup>

Resistance of *Candida albicans* is due to its ability to increase expression or decrease susceptibility of the target, spontaneous mutations, and escape host immune defenses.<sup>58</sup> Zaidi et al, indicated that the evolutionary changes in the total protein profile enhanced the resistance of *Candida albicans*.<sup>59</sup>

This study showed that toothpaste containing gambir had antibacterial and antifungal activities against *Streptococcus mutans*, *Porphyromonas gingivalis*, and *Candida albicans*. So that it also showed that *Uncaria gambir* plants, which are widely distributed in South Sumatera and are traditionally used to treat a variety of diseases, were effective in reducing bacteria and candida in the oral cavity.

## CONCLUSION

It concluded that toothpaste containing gambir (*Uncaria gambir*) has antimicrobial potency.

**Acknowledgement:** The authors wish to gratefully acknowledge the financial support from Universitas Sriwijaya.

**Author Contributions:** Conceptualization, SRPD; Methodology, SRPD and PH; software, STM; validation, SRPD, PH, DA and STM.; formal analysis STM; investigation, SRPD and STM; resources, SRPD and STM; data curation, STM; writing original draft preparation, SRPD; writing review and editing, PH; visualization, DA, supervision, SRPD and PH; project administration, STM.; funding acquisition, SRPD and STM. All authors have read and agreed to the published version of the manuscript."

**Funding:** This research received only from Universitas Sriwijaya.

**Institutional Review Board Statement:** Ethical review and approval were waived for this study due to "Not applicable" for studies not involving humans or animals.

**Data Availability Statement:** Data supporting reported results can be found, including links to publicly archived datasets analyzed or generated during the study.

**Conflicts of Interest:** The authors declare no conflict of interest.

## REFERENCES

1. Kementerian Kesehatan Republik Indonesia. Riset Kesehatan Dasar (Riskesdas) 2018. Jakarta: Kementerian Kesehatan RI; 2018, pp 114-118.
2. Rathee M, Sapra A. Dental Caries. [Updated 2022 Jun 12]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK551699/>
3. Lemos JA, Palmer SR, Zeng L, Wen ZT, Kajfasz JK, Freires IA, Abranches J, Brady LJ. The Biology of *Streptococcus mutans*. Microbiol Spectr. 2019 Jan; 7(1): 10. doi: [10.1128/microbiolspec.GPP3-0051-2018](https://doi.org/10.1128/microbiolspec.GPP3-0051-2018).
4. Zhang Y, Fang J, Yang J, Gao X, Dong L, Zheng X, Sun L, Xia B, Zhao N, Ma Z, Wang Y. *Streptococcus mutans*-associated bacteria in dental plaque of severe early childhood caries. Journal of Oral Microbiology 2022; 14: 1. DOI: [10.1080/20002297.2022.2046309](https://doi.org/10.1080/20002297.2022.2046309)
5. Könönen E, Gursoy M, Gursoy UK. Periodontitis: A Multifaceted Disease of Tooth-Supporting Tissues. J Clin Med. 2019 Jul 31; 8(8):1135. DOI: [10.3390/jcm8081135](https://doi.org/10.3390/jcm8081135).
6. How KY, Song KP, Chan KG. *Porphyromonas gingivalis*: An Overview of Periodontopathogenic Pathogen below the Gum Line. Front Microbiol. 2016 Feb 9;7:53. DOI: [10.3389/fmicb.2016.00053](https://doi.org/10.3389/fmicb.2016.00053)

7. Mysak J, Podzimek S, Sommerova P, Lyuya-Mi Y, Bartova J, Janatova T, Prochazkova J, Duskova J. *Porphyromonas gingivalis*: major periodontopathic pathogen overview. J Immunol Res. 2014;2014:476068. DOI: [10.1155/2014/476068](https://doi.org/10.1155/2014/476068).
8. Stephen J. Challacombe, Penelope J. Shirlaw, Martin H. Thornhill. Chapter 102 - Immunology of Diseases of the Oral Cavity. Editor(s): Mestecky J, Strobel MW, Kelsall BL, Cheroutre H, Lambrecht BN. Mucosal Immunology (Fourth Edition). Academic Press: 2015. 1943-1983. DOI: [10.1016/B978-012491543-5/50093-0](https://doi.org/10.1016/B978-012491543-5/50093-0)
9. Vila T, Sultan AS, Montelongo-Jauregui D, Jabra-Rizk MA. Oral Candidiasis: A Disease of Opportunity. J Fungi (Basel). 2020 Jan 16;6(1):15. DOI : [10.3390/jof6010015](https://doi.org/10.3390/jof6010015).
10. Rao PS, Majumdar RS, Anil S. Clinical Appearance of Oral Candida Infection and Therapeutic Strategies. Frontiers in Microbiology 2015; 6. DOI: [10.3389/fmicb.2015.01391](https://doi.org/10.3389/fmicb.2015.01391)
11. Vyas T, Bhatt G, Gaur A, Sharma C, Sharma A, Nagi R. Chemical plaque control - A brief review. J Family Med Prim Care. 2021 Apr;10(4):1562-1568. DOI: [10.4103/ijfmpc](https://doi.org/10.4103/ijfmpc).
12. Nwakanma C, Ejim JN, Unachukwu M. The Effects of selected toothpaste on the microbial flora of the mouth of GOU Student. Int J Curr Microbiol Appl Sci. 2014; 3(9): pp 785-92
13. Manja NK. Review on- ingredients used in toothpaste formulation. Indian J of Medical Research and Pharmaceutical Sciences 2020; 7(9): pp 9-15.
14. Thornton-Evans G, Junger ML, Lin M, Wei L, Espinoza L, Beltran-Aguilar E. Use of Toothpaste and Toothbrushing Patterns Among Children and Adolescents - United States, 2013-2016. MMWR Morb Mortal Wkly Rep. 2019 Feb 1;68(4): pp 87-90. DOI: [10.15585/mmwr.mm6804a3](https://doi.org/10.15585/mmwr.mm6804a3).
15. Septiani S, Indrawan D, Arista GS, Rakhmat A, Sari YW, Nuzulia NA, Wahyuni WT. Choosing herbal toothpaste: study on consumer behaviour and preferences in the greater Jakarta area. Ju Aplikasi Manajemen and Bisnis 2022; 8(3): pp 758-68. DOI: [10.17358/jabm.8.3.758](https://doi.org/10.17358/jabm.8.3.758)
16. Hidayaturrehman, Asidiki H, Sari CN, Harismah N. Development of Herbal Toothpaste Formulation with Combination of Binahong and Stevia (Stevia Rebaudina) Leaves Extract and Lemon Juice. J Nutraceutical and Herbal Med. 2020; 3(1): pp 15-22
17. Nuraskin CA, Addilla ID, Reza. Effectiveness Of Herbal And Non-Herbal Toothpaste To Decrease Plac Index In Class Iv Students At Sd Neg E Ri 62 Cot Mesjid, Banda Aceh. Dental Health J of Aceh 2022; 1(2): 34-9. DOI: [10.30867/dheja.v1i2.110](https://doi.org/10.30867/dheja.v1i2.110)
18. Yaman E, Woerdenbag HJ, Kayser O. Jamu: Indonesian traditional herbal medicine towards rational phytopharmacological use. J Herbal Med 2014; 4(2): 34-41. DOI: [10.1016/j.hermed.2014.01.002](https://doi.org/10.1016/j.hermed.2014.01.002)
19. Auliana FR, Ifora I, Fauziah F. Phytochemical and Anti-Inflammatory of *Uncaria gambir*: A Review Asian J of Pharmaceutical Research and Development. 2022; 10(1): pp. 79-83 DOI : [10.22270/ajpr.v10i1.1077](https://doi.org/10.22270/ajpr.v10i1.1077)
20. Dewi SRP, Marlamsya DO, Bikarindrasari R. Efek antikaries ekstrak gambir pada tikus jantan galur Wistar. Maj. Ked. Gigi Indonesia 2017; 3(2): 83-92. DOI: [10.22146/majkedgiind.17407](https://doi.org/10.22146/majkedgiind.17407)
21. Wu M, Brown AC. Applications of Catechins in the Treatment of Bacterial Infections. Pathogens. 2021 May 1;10(5): pp 546. DOI: [10.3390/pathogens10050546](https://doi.org/10.3390/pathogens10050546).
22. Melia S, Novia D, Juliyarsi I. Antioxidant and Antimicrobial Activities of Gambir (*Uncaria gambir* Roxb) Extracts and Their Application in Rendang. Pakistan Journal of Nutrition 2015; 14(12):938-41. DOI: [10.3923/pjn.2015.938.941](https://doi.org/10.3923/pjn.2015.938.941)
23. Saito H, Tamura M, Imai K, Ishigami T, Ochiai K. Catechin inhibits *Candida albicans* dimorphism by disrupting Cek1 phosphorylation and cAMP synthesis. Microb Pathog. 2013 Mar;56:16-20. DOI: [10.1016/j.micpath.2013.01.002](https://doi.org/10.1016/j.micpath.2013.01.002).
24. Ifora I, Efelzita D, Bellatasie R, Uyun HSK. Antifungal Potential of Purified Gambier (*Uncaria gambir* Roxb.). Int J Pharm Sci Med. 2023; 8 (2): pp. 40-43. DOI: [10.47760/ijpsm.2023.v08i02.004](https://doi.org/10.47760/ijpsm.2023.v08i02.004)
25. Dewi SRP, Kamaluddin MT, Theodorus, Pambayun R. Anticariogenic effect of gambir (*Uncaria Gambir* [Roxb.]) extract on enamel tooth surface exposed by *Streptococcus mutans*. Int. J Health Sci. Res. 2016; 6(8): pp 171-9
26. Isromarina R, Rosa E, Rusli D. Antibacterial activity of *Uncaria gambir* (Hunter) Roxb extract leaves against *Vibrio Cholerae* ATCC 14033. Jurnal Ilmiah Bakti Farmasi 2019; 4(1): pp 21-6
27. Dewi SRP, Pratiwi A, Teodorus. The effect of gambir extract (*Uncaria gambir* [Roxb]) as antiseptic on gingival wound in rats. Odonto J. 2018; 5(1): pp 80-8
28. Aspinall SR, Parker JK, Khutoryanskiy VV. Oral care product formulations, properties and challenges. Colloids Surf B Biointerfaces. 2021 Apr; 200: pp 111567. DOI: [10.1016/j.colsurfb.2021.111567](https://doi.org/10.1016/j.colsurfb.2021.111567).
29. Uc-Cachón AH, Gracida-Osorno C, Luna-Chi IG, Jiménez-Guillermo JG, Molina-Salinas GM. High Prevalence of Antimicrobial Resistance Among Gram-Negative Isolated Bacilli in Intensive Care Units at a Tertiary-Care Hospital in Yucatán Mexico. Medicina (Kaunas). 2019 Sep 13; 55(9): pp 588. DOI: [10.3390/medicina55090588](https://doi.org/10.3390/medicina55090588).
30. de Vries C, Ruacho G, Kindstedt E, Potempa BA, Potempa J, Klinge B, Lundberg P, Svenungsson E, Lundberg K. Antibodies to *Porphyromonas gingivalis* Are Increased in Patients with Severe Periodontitis, and Associate with Presence of Specific Autoantibodies and Myocardial Infarction. J Clin Med. 2022; 11(4): pp. 1008. DOI: [10.3390/jcm11041008](https://doi.org/10.3390/jcm11041008)
31. Smithee S, Tracy S, Drescher KM, Pitz LA, McDonald T. A novel, broadly applicable approach to isolation of fungi in diverse growth media. Journal of Microbiological Methods 2014; 105: 155-61. DOI: [10.1016/j.mimet.2014.07.023](https://doi.org/10.1016/j.mimet.2014.07.023).
32. Nafisa S, Swandiny GF, Gangga E, Zaenudin YA. Antimicrobial activity and phytochemical screening of *Citrus aurantifolia* leaves ethanolic extract. Jurnal Ilmu Kefarmasian Indonesia 2021; 19(2): 287-91. DOI: [10.20959/wjpr20171-7634](https://doi.org/10.20959/wjpr20171-7634)
33. Marlinda. Identifikasi kadar katekin pada gambir (*Uncaria gambier* Roxb.). Jurnal Optimalisasi. 2018; 4(1): 47-53 DOI: [10.35308/jopt.v4i1.1474](https://doi.org/10.35308/jopt.v4i1.1474)
34. Inmawaty J, Indrati, Satari MH. Inhibitory concentrations of gambir (*Uncaria gambir* Roxb.) catechins extract against *Streptococcus mutans*. Padjajaran J Dent. 2012; 24(3): 161-6 DOI : [10.24198/pjd.vol24no3.26832](https://doi.org/10.24198/pjd.vol24no3.26832)
35. Bernatoniene J, Kopustinskiene DM. The Role of Catechins in Cellular Responses to Oxidative Stress. Molecules. 2018; 23(4): 965. DOI: [10.3390/molecules23040965](https://doi.org/10.3390/molecules23040965).
36. Wu M, Brown AC. Applications of Catechins in the Treatment of Bacterial Infections. Pathogens. 2021 May 1;10(5):546. DOI: [10.3390/pathogens10050546](https://doi.org/10.3390/pathogens10050546).
37. Mai-Prochnow A, Clauson M, Hong J, Murphy AB. Gram positive and Gram negative bacteria differ in their sensitivity to cold plasma. Sci Rep. 2016 Dec 9;6:38610. DOI: [10.1038/srep38610](https://doi.org/10.1038/srep38610).
38. Taylor PW. Interactions of Tea-Derived Catechin Gallates with Bacterial Pathogens. Molecules. 2020; 25(8):1986. DOI :[10.3390/molecules25081986](https://doi.org/10.3390/molecules25081986)
39. Ren Z, Chen L, Li J, Li Y. Inhibition of *Streptococcus mutans* polysaccharide synthesis by molecules targeting glycosyltransferase activity. J Oral Microbiol 2016; 18(1): 31095. DOI : [10.3402/jom.v8.31095](https://doi.org/10.3402/jom.v8.31095)
40. Xu X, Zhou XD, Wu CD. The tea catechin epigallocatechin gallate suppresses cariogenic virulence factors of *Streptococcus mutans*. Antimicrob Agents Chemother. 2011; 55(3): 1229-36
41. Kong C, Zhang H, Li L. Effects of green tea extract epigallocatechin-3-gallate (EGCG) on oral disease-associated microbes: a review. J Microbiol. 2022; 14(1): 1-14. DOI: [10.1080/20002297.2022.2131117](https://doi.org/10.1080/20002297.2022.2131117)
42. Lestari C. The effect of subgingival irrigation of gambir (*Uncaria gambir* [Hunter] Roxb) catechin to the number of *Actinobacillus actinomycetemcomitans* in the gingival sulcus of periodontitis mice. Padjajaran J Dent. 2011; 23(3): 167-71. DOI: [10.24198/pjd.vol23no3.14033](https://doi.org/10.24198/pjd.vol23no3.14033)

43. Sari LA, Deynilisa S. Effectiveness of gargling gambier sap boiled water for the treatment of gingivitis. *J Kesehatan Gigi dan Mulut* 2019; 1(2): 17-20.
44. Kurhekar JV. Tannins-antimicrobial chemical components. *Int J Tec Sci*. 2016; 9: p 5-9
45. Kaczmarek B. Tannic Acid with Antiviral and Antibacterial Activity as A Promising Component of Biomaterials-A Minireview. *Materials* (Basel). 2020 Jul 20;13(14): pp 3224. DOI: [10.3390/ma13143224](https://doi.org/10.3390/ma13143224).
46. Ho KY, Tsai CC, Huang JS, Chen CP, Lin TC, Lin CC. Antimicrobial activity of tannin components from *Vaccinium vitis-idaea* L. *Journal of Pharmacy and Pharmacology*. 2001; 53(2): 187-91.
47. Choi O, Yahiro K, Morinaga N, Miyazaki M, Noda M. Inhibitory effects of various plant polyphenols on the toxicity of *staphylococcal*-toxin. *J Microb Pathog*. 2007; 42: 215-24.
48. Aboody MSA, Mickymaray S. Anti-Fungal Efficacy and Mechanisms of Flavonoids. *Antibiotics* (Basel). 2020; 9(2): 45. DOI: [10.3390/antibiotics9020045](https://doi.org/10.3390/antibiotics9020045).
49. Anand J, Rai N. Anticandidal synergistic activity of green tea catechins, antimycotics and copper sulphate as a mean of combinational drug therapy against candidiasis. *J Mycol Med*. 2017; 27(1): 33-45. DOI: [10.1016/j.mycmed.2016.08.004](https://doi.org/10.1016/j.mycmed.2016.08.004).
50. Congyi Zhu, Mengying Lei, Mebeaselassie Andargie, Jiwu Zeng, Jianxiong Li. Antifungal activity and mechanism of action of tannic acid against *Penicillium digitatum*. *Physiological and Molecular Plant Pathology* 2019; 107: 46-50. DOI: [10.1016/j.pmpp.2019.04.009](https://doi.org/10.1016/j.pmpp.2019.04.009).
51. Rahayuningsih E, Setiawan FA, Ayanie CJ, Aditya A, Ayuningtyas YI. A Study on the Effectiveness of Antifungal Agents on Cotton Fabrics Colored by Gambir Natural Dyes (*Uncaria gambir*). *J Konversi Universitas Muhammadiyah Jakarta* 2019; 8(1): 45-54. DOI: [10.24853/konversi.8.1.10](https://doi.org/10.24853/konversi.8.1.10).
52. Johnston NR, Cline G, Strobel SA. Cells Adapt to Resist Fluoride through Metabolic Deactivation and Intracellular Acidification. *Chem Res Toxicol*. 2022; 35(11): 2085-96. DOI: [10.1021/acs.chemrestox.2c00222](https://doi.org/10.1021/acs.chemrestox.2c00222).
53. Sperandio FF, Huang YY, Hamblin MR. Antimicrobial photodynamic therapy to kill Gram-negative bacteria. *Recent Pat Antiinfect Drug Discov*. 2013; 8(2): 108-20. DOI: [10.2174/1574891x113089990012](https://doi.org/10.2174/1574891x113089990012).
54. Budisa N, Kubyskhin V, Schulze-Makuch D. Fluorine-rich planetary environments as possible habitats for life. *Life* (Basel). 2014; 4(3): 374-85. DOI: [10.3390/life4030374](https://doi.org/10.3390/life4030374).
55. Li S, Breaker RR. Fluoride enhances the activity of fungicides that destabilize cell membranes. *Bioorg Med Chem Lett*. 2012; 22(9): 3317-22. DOI: [10.1016/j.bmcl.2012.03.006](https://doi.org/10.1016/j.bmcl.2012.03.006).
56. Mozayeni MA, Hadian A, Bakhshaei P, Dianat O. Comparison of Antifungal Activity of 2% Chlorhexidine, Calcium Hydroxide, and Nanosilver gels against *Candida Albicans*. *J Dent* (Tehran). 2015; 12(2): 109-17.
57. Adwan G, Salameh Y, Adwan K, Barakat A. Assessment of antifungal activity of herbal and conventional toothpastes against clinical isolates of *Candida albicans*. *Asian Pac J Trop Biomed*. 2012; 2(5): 375-9. DOI: [10.1016/S2221-1691\(12\)60059-8](https://doi.org/10.1016/S2221-1691(12)60059-8).
58. Costa-de-Oliveira S, Rodrigues AG. *Candida albicans* Antifungal Resistance and Tolerance in Bloodstream Infections: The Triad Yeast-Host-Antifungal. *Microorganisms*. 2020 Jan 22;8(2):154. DOI: [10.3390/microorganisms8020154](https://doi.org/10.3390/microorganisms8020154).
59. Zaidi KU, Mani A, Thawani V, Mehra A. Total Protein Profile and Drug Resistance in *Candida albicans* Isolated from Clinical Samples. *Mol Biol Int*. 2016;2016:4982131. DOI: [10.1155/2016/4982131](https://doi.org/10.1155/2016/4982131).